AMENDMENTS TO THE CLAIMS (AS ON AMENDED SHEETS ANNEXED TO IPER)

This listing of claims will replace all prior versions, and listings, of claims in the application:

1. (original) Process for the biochemical synthesis of 6-amino caproic acid wherein either 6-aminohex-2-enoic acid of formula [1] (6-AHEA)

$$H_2N - CH_2 - CH_2 - CH_2 - CH = CH - COOH$$
 [1]

or wherein 6-amino-2-hydroxy-hexanoic acid (6-AHHA), a compound capable of being transformed into 6-aminohex-2-enoic acid, is treated with an enzyme having α,β -enoate reductase activity towards molecules containing an α,β -enoate group and a primary amino group, in particular with an enzyme having α,β -enoate reductase activity towards 6-aminohex-2-enoic acid.

- 2. (original) Process according to claim 1, characterized in that the enzyme having α,β-enoate reductase activity is an enzyme originating from a microorganism from the group of species of *Acetobacterium* sp., *Acremonium* sp., *Agrobacterium* sp., *Burkholderia* sp., *Cephalosporium* sp., *Clostridium* sp., *Escherichia* sp., *Moorella* sp., *Ochrobactrum* sp., *Pseudomonas* sp., *Salmonella* sp., *Shigella* sp., *Tilachlidium* sp., *Yersinia* sp., and *Vibrio* sp.
- 3. (currently amended) Process according to one of claims 1 or 2, characterized in that the enzyme having α,β-enoate reductase activity is an enzyme originating from *Acremonium* sp., *Clostridium* sp., *Moorella* sp. or *Ochrobactrum* sp.
- 4. (original) Process according to claim 3, characterized in that the enzyme having is an enzyme from *Acremonium strictum* CBS114157, *Clostridium tyrobutyricum* DSM1460, *Moorella thermoacetica* DSM1974, *Ochrobactrum anthropi* NCIMB41200, or *Clostridium kluyveri* DSM555.

- 5. (currently amended) Process according to claim 1 or 2, characterized in that the enzyme having α,β-enoate reductase activity has aerostable α,β-enoate reductase activity and is an enzyme originating from a microorganism from the group of species of *Agrobacterium* sp., *Burkholderia* sp., *Escherichia* sp., *Pseudomonas* sp., *Salmonella* sp., *Shigella* sp., *Yersinia* sp., and *Vibrio* sp.
- 6. (original) Process according to claim 5, characterized in that the enzyme having aerostable α,β-enoate reductase activity is an enzyme originating from an *Escherichia coli* species.
- 7. (original) Process according to claim 6, characterized in that the enzyme having aerostable α,β-enoate reductase activity is an enzyme originating from from Escherichia coli K12.
- 8. (currently amended) Process according to any of claims 1-7 claim 1, characterized in that 6-aminohex-2-enoic acid is being converted into 6-amino caproic acid at a pH in the range of from 3 to 9.
- 9. (original) Process according to claim 8, characterized in that, the pH is in the range of from 4 to 8.
- 10. (original) Process according to claim 9, characterized in that the pH is in the range of from 5 to 8.
- 11. (original) Process according to claim 8, characterized in that, the pH is in the range of from 5.5 to 7 under anaerobic conditions and of from 6.5 to 8 under aerobic conditions.
- 12. (currently amended) Process according to any of claims 1-11 claim 1, characterized in that the process is carried out in a host organism selected from the group of genera consisting of Aspergillus, Bacillus, Corynebacterium, Escherichia and Pichia.

- 13. (original) Process according to claim 12, characterized in that the process is carried out in a host organism selected from the group of *Escherichia coli*, *Bacillus*, *Corynebacterium glutamicum*, *Aspergillus niger* or *Pichia pastoris* host organisms.
- 14. (currently amended) Process according to claim 12 or 13, characterized in that in the host organism an α,β-enoate reductase gene encoding an enzyme having α,β-enoate reductase activity towards molecules containing an α,β-enoate group and a primary amino group is cloned and expressed.
- 15. (original) An *Escherichia coli* host cell wherein the α,β-enoate reductase gene from *Ochrobactrum anthropi* NCIMB41200, or from *Acremonium strictum* CBS114157 is cloned and expressed.
- 16. (original) A Bacillus host cell wherein the α,β-enoate reductase gene from Moorella thermoacetica DSM1974, or from Clostridium tyrobutyricum DSM1460, or from Ochrobactrum anthropi NCIMB41200, or from Acremonium strictum CBS114157 is cloned and expressed.
- 17. (original) A *Corynebacterium glutamicum* host cell wherein the α,β-enoate reductase gene from *Moorella thermoacetica* DSM1974, or from *Clostridium tyrobutyricum* DSM1460, or from *Ochrobactrum anthropi* NCIMB41200, or from *Acremonium strictum* CBS114157 is cloned and expressed.
- 18. (original) An Aspergillus niger host cell wherein the α,β-enoate reductase gene from Acremonium strictum CBS114157, or from Moorella thermoacetica DSM1974, or from Clostridium tyrobutyricum DSM1460, or from Ochrobactrum anthropi NCIMB41200 is cloned and expressed.
- 19. (original) A *Pichia pastoris* host cell wherein the α,β-enoate reductase gene from *Acremonium strictum* CBS114157, or from *Moorella thermoacetica*

- DSM1974, or from *Clostridium tyrobutyricum* DSM1460, or from *Ochrobactrum anthropi* NCIMB41200 is cloned and expressed.
- 20. (original) A host cell selected from the group of *Aspergillus*, *Bacillus*, *Corynebacterium*, and *Pichia* host cells, in which the aerostable α,β-enoate reductase gene *nemA* from *E. coli* K12 is cloned and expressed.
- 21. (original) Process for precursor fermentation of 6-amino caproic acid starting either from 6-aminohex-2-enoic acid (6-AHEA) or from 6-amino-2-hydroxyhexanoic acid (6-AHHA), and applying at least an enzymatic step with an enzyme having α,β-enoate reductase activity towards molecules containing an α,β-enoate group and a primary amino group, in particular with an enzyme having α,β-enoate reductase activity towards 6-aminohex-2-enoic acid.
- 22. (original) Process according to claim 21, characterized in that the process is performed in a microorganism wherein 6-aminohex-2-enoic acid is being formed in vivo.
- 23. (original) Process according to claim 22, characterized in that6-aminohex-2-enoic acid is being formed *in vivo* from solutions or slurriescontaining a suitable carbon source.
- 24. (original) Biochemically produced 6-aminohex-2-enoic acid, having a ¹²C versus ¹³C versus ¹⁴C isotope ratio of about the same value as occurring in environmental carbon dioxide.
- 25. (original) Biochemically produced 6-amino-hexanoic acid having a ¹²C versus ¹³C versus ¹⁴C isotope ratio of about the same value as occurring in environmental carbon dioxide.
- 26. (original) ε-Caprolactam produced from biochemically produced
 6-aminohex-2-enoic acid or 6-amino-hexanoic acid, and having a ¹²C versus ¹³C

- versus ¹⁴C isotope ratio of about the same value as occurring in environmental carbon dioxide.
- 27. (original) Nylon-6 and other derivatives produced from any of the biochemically produced products of claims 24 or 25, or from ε-caprolactam according to claim 26, and having a ¹²C versus ¹³C versus ¹⁴C isotope ratio of about the same value as occurring in environmental carbon dioxide.